| NAME OF THE COURSE Molecular Cell Biology  |   |        |  |           |    |    |   |  |
|--|---|--------|--|-----------|----|----|---|--|
| Code   | PMB701  |        | Year of study                            | 1.        |    |    |   |  |
| Course teacher   | Ivana Novak Nakir, Ph.D,<br>Associate Professor<br>Ivica Šamanić, Ph.D.,<br>Assistant Professor<br>Katarina Trajković, Ph.D,<br>Research Assistant  |        | Credits (ECTS)                           | 8         |    |    |   |  |
| Associate teachers   | Mija Marinković, Ph.D   |        | Type of instruction<br>(number of hours) | L         | S  | E  | F |  |
| Status of the course   | Obligatory  |        | Percentage of application of e-learning  | 45<br>10% | 15 | 45 |   |  |
|  |   | COURSE | E DESCRIPTION                            |           |    |    |   |  |
| Course objectives<br>Course enrolment<br>requirements and<br>entry competences<br>required for the<br>course | Understanding of the basic cell structures and regulatory cell biology mechanisms<br>on a molecular level. Students will be introduced to current molecular and cellular<br>biology research procedures in a hands-on environment. This course develops<br>technical skills and prepare students for a career in science or research work.<br>Recommended fundamental knowledge of cell biology, genetics, biochemistry and<br>molecular biology.<br>After successful completion of this course students will be able to:<br>• links the structure and function of the cellular organelles to the mechanisms that<br>modulate cellular response to external or internal stimuli<br>• use equipment and reagents properly, maintaining a safe laboratory environment   |        |  |           |    |    |   |  |
| expected at the<br>level of the course<br>(4 to 10 learning<br>outcomes)                                     | <ul> <li>discuss research results in the context of the scientific literature through written<br/>and oral communications</li> <li>describe and carry out basic cell culture and microscopy</li> <li>write a lab report that includes an evaluation of the results obtained in the<br/>laboratory</li> <li>present research in a seminar form</li> <li>work cooperatively in a molecular biology lab</li> </ul>   |        |  |           |    |    |   |  |
| Course content<br>broken down in<br>detail by weekly<br>class schedule<br>(syllabus)                         | LECTURES:<br>INTRODUCTION-Structural and functional organization of cells, The chemical<br>components of a cell<br>NUCLEAR STRUCTURE AND DYNAMICS - Nuclear matrix or nuclear skeleton,<br>Nuclear Bodies (nucleolus - RNA Synthetic Apparatus, Cajal bodies, promyelocytic<br>leukemia nuclear bodies, PML-NBs), nuclear transport (mRNA transport),<br>Chromatin structure and function<br>FUNDAMENTAL MOLECULAR GENETIC MECHANISMS – From DNA to protein<br>(the Central dogma), The ribosome (the ribosome is a Ribozyme), Regulated<br>synthesis and degradation of proteins (ubiquitin and proteasome – mediated<br>proteolysis)<br>THE CELL CYCLE - S phase, Mitosis and Cytokinesis, Control of the Cell Cycle:<br>The Role of Protein Kinases (regulation of CDK activity), Checkpoints, Cdk<br>Inhibitors; Genetic Control of Meiosis, Deregulation of the Cell cycle |        |  |           |    |    |   |  |

| THE STRUCTURE AND FUNCTION OF THE PLASMA MEMBRANE – The Lipid   |
|---|
| Bilayer: Composition and Structural Organisation; Membrane Proteins: Structure  |
| and Basic Functions   |
| CYTOPLASMATIC MEMBRANE SYSTEMS – The Endoplasmatic Reticulum,   |
| The Golgi complex, Types of Vesicle Transport, Moving Proteins into Membranes   |
| and Organelles, Glycosylation, Vesicular Traffic, Secretion, and Endocytosis,   |
| Autophagy   |
| CELLULAR BIOENERGETICS AND METABOLISM – Catalysis and the energy  |
| consumption (oxidation and reduction, carrier molecules – ATP, NADH and   |
| NADPH), The structure and functions of Mitochondria (ATP production in  |
| mitochondria), Chloroplast and photosynthesis (molecular analysis of  |
| photosystems)   |
| INTERACTION BETWEEN CELLS AND THEIR ENVIRONMENT - Cell  |
| Junctions, Cell Adhesion (Cell adhesion molecules - transmembrane proteins), and  |
|   |
| the Extracellular Matrix (Proteoglycans, Fibrous proteins - collagen and elastin)                                       |
| THE CYTOSKELETON AND CELL MOTILITY- Structure and Function of   |
| Microtubules, Intermediate Filaments, Actin, Myosin, Cellular Motility  |
| CELLULAR RESPONSES TO DNA DAMAGE – DNA damage detection and   |
| signaling (Protein-Tyrosine Phosphorylation as a Mechanism for Signal   |
| Transduction), Transcriptional Response (Post-transcriptional gene control), DNA  |
| Damage Checkpoints, DNA Repair Mechanisms, Apoptosis (Programmed Cell   |
| Death) and non-apoptotic types of cell death (necroptosis, pyroptosis, anoikis,   |
| mitotic catastrophe)  |
| CELL SIGNALING AND SIGNAL TRANSDUCTION – The Basic Elements of  |
| Cell Signaling, Signal Transduction by G Protein-coupled receptors, The Ras-Map   |
| Kinase Pathway, Signaling by the Insulin Receptor, The roles of Calcium and NO as                                       |
| an Intracellular Messengers, Cytokine receptors and the JAK/STAT signaling  |
| pathway, Signaling pathways: Wnt and Notch, Hedgehog and NF-ĸB,   |
| BIOLOGY OF SPECIFIC CELL TYPES AND CELL DIFFERENTIATION – Cell  |
| reprogramming and pluripotent stem cells, Nerve cells (neurons), Immune cells   |
| (Natural killer cells, B-cels, T-cells), Adipose tissue (adipocytes), Cell memory,                                      |
| Mechanisms of cell polarity and asymmetric cell division, Cancer cells.   |
| LABORATORY:   |
| Students will spend 4h at MedILS, where they will be introduced to laboratory work                                      |
| related to cell biology – cell culture room, main laboratory, microscopes. Samples                                      |
| containing cells with fluorescently labeled organelles will be observed under the                                       |
| fluorescent microscope. At the end, students will write a laboratory report which will                                  |
| be evaluated by the lecturer.   |
| Experimental Topic.   |
| <ul> <li>microscopy methods for structural analysis of the eukaryotic cell</li> </ul>                                   |
| - cultivation of eukaryotic cells   |
| <ul> <li>organelle isolation</li> <li>cell cycle analysis by quantitation of DNA content with flow cytometry</li> </ul> |
| <ul> <li>analysis of cell viability using Propidium Iodide</li> </ul>   |
| - DNA damage sensitivity - In vitro non-homologous DNA end joining assays   |
| - Spatio-temporal dynamics of protein   |
| assembly/disassembly at sites of DNA breaks   |
| - expression and localization of a fusion protein of a targeted gene and a  |
| reporter gene   |
| <ul> <li>monitoring of autophagy by fluorescent labeling of autophagosomal</li> </ul>                                   |
| vesicles  |

|  | <ul> <li>Isolation and purification of receptor proteins by affinity chromatography</li> <li>Co-Immunoprecipitation: Isolation of protein signaling complexes from native tissues</li> <li>SEMINARS:</li> <li>Reading and discussing primary scientific literature, writing a short assay summarizing analyzed articles. Selected articles related to the above topics will be analyzed in 2-hour blocks. Assays will be written at home and submitted to the lecturer for evaluation.</li> </ul>   |           |                  |  |                                       |                                 |
|--|---|-----------|------------------|--|---------------------------------------|---------------------------------|
| Format of<br>instruction   | <ul> <li>☑ lectures</li> <li>☑ seminars and workshops</li> <li>☑ exercises</li> <li>□ on line in entirety</li> <li>☑ partial e-learning</li> <li>□ field work</li> </ul>  |           |                  | <ul> <li>☑ independent assignments</li> <li>☑ multimedia</li> <li>☑ laboratory</li> <li>☑ work with mentor</li> <li>□ (other)</li> </ul> |                                       |                                 |
| Student<br>responsibilities  | Attendance at all lectures is strongly recommended.<br>Students are required to attend laboratory exercises and write lab reports.  |           |                  |  |                                       |                                 |
| Screening student<br>work (name the<br>proportion of ECTS<br>credits for each<br>activity so that the<br>total number of<br>ECTS credits is<br>equal to the ECTS | Class<br>attendance   | 1         | Research         |  | Practical trainir                     | ng 1                            |
|  | Experimental<br>work  |           | Report           |  | Lab reports                           | 1                               |
|  | Essay   |           | Seminar<br>essay | 1  | Quizzes                               | 1                               |
|  | Tests   |           | Oral exam        |  | (Other)                               |                                 |
| value of the course)   | Written exam  | 3         | Project          |  | (Other)                               |                                 |
| Grading and<br>evaluating student<br>work in class and at<br>the final exam  | <ul> <li>Methods of Evaluation</li> <li>Lab reports</li> <li>All lab reports must contain complete and detailed outline of the experimental procedure, description of the results accompanied by analysis and interpretation.</li> <li>Quizzes</li> <li>Quizzes will cover both material from the previous labs as well as material on the experiment for the present lab session (based on theory and protocols found in the lab manual and interpretation of data)</li> <li>Practical skills</li> <li>At times, students will be asked to demonstrate techniques of accurate preparation of chemical solution and reagents, correct assembly of reactions according to protocol, ability to pipette correctly, competent use of standard and specialized equipment and sterile technique.</li> <li>Research-based class seminar will be elevated.</li> <li>Students will have to prepare presentation showing background of the problem they are dealing with. The presentation will be scored according to the content of the presentation (key words, critical review of literature, presentation of scientific results), format, innovativeness and language competence as well.</li> <li>Class Participation will also be part of the grade.</li> <li>Final Lecture Exam: written examination.</li> </ul> |           |                  |  |                                       |                                 |
| Required literature<br>(available in the<br>library and via other  | Title   |           |                  |  | Number of<br>copies in<br>the library | Availability via<br>other media |
| media)   | 1. Lodish, Harv   | ey - Mole | cular cell biol  | ogy  |                                       |                                 |

|  | 8th edition.: New York, NY: W. H. Freeman, 2016.   |
|--|--|
| Optional literature<br>(at the time of<br>submission of study<br>programme | Gerald Karp, Janet Iwasa, Wallace Marshall - Karp's Cell and Molecular Biology-<br>Wiley E-Text, 2016<br>Lab Manual: Lab exercises will be provided in the form of handouts. |
| proposal)<br>Quality assurance<br>methods that<br>ensure the               | Student evaluation   |
| acquisition of exit<br>competences<br>Other (as the                        |  |
| proposer wishes to<br>add)   |  |