NAME OF THE COURSE Cytogenetic Chromosome Analysis							
Code	PPB25	3	Year of study	3			
Course teacher	Assista Ivica Ša	nt Professor amanić, PhD	Credits (ECTS)	2			
Associate teachers	Professor Jasna Puizina, PhD Assistant Professor Željana Fredotović, PhD		Type of instruction (number of hours)	L 10	S 5	E 15	F
Status of the course	Elective	9	Percentage of application of e-learning	10%			
		COURSE	DESCRIPTION	•			
Course objectives	Insight into the molecular and structural dynamics of mitotic and meiotic chromosomes. Theoretical and practical introduction of students with the classical and molecular cytogenetic techniques.						
Course enrolment requirements and entry competences required for the course	None	· · · · ·					
Learning outcomes expected at the level of the course (4 to 10 learning outcomes)	 Student will be able to: The integration and implementation of all the knowledge acquired during the various courses (primarily Cell biology, Genetics and Molecular biology) for studying genomes at the level of chromosomes and chromatin. Explain the importance of cytogenetics in the area of basic research as well as its applications in medical genetics, biotechnology and agriculture The skills and knowledge acquired throughout the training will enable students to perform in situ hybridization and other molecular techniques needed to work in the Molecular and Cytogenetic laboratories (employment of cytogenetic technologists or clinical laboratory technicians). The acquired knowledge and skills will form the bases for further research 						
Course content broken down in detail by weekly class schedule (syllabus)	Lectures: 1. CYTOGENETICS METHODS: Molecular cytogenetic techniques; In situ hybridization (FISH, GISH, direct visual <i>in situ</i> hybridization (DIRVISH) on elongated DNA fibers), <i>in situ</i> PCR, PRINS (PRimed <i>IN Situ</i> labeling), Flow cytometry, Chromosome microdissection. Classical cytogenetic techniques; chromosome preparations, karyotyping, G-(Giemsa), R-(reverse), C-(centromere) and Q-(quinacrine) banding, chromosome labeling. 2. CHROMATIN STRUCTURE: Histones, DNA, nucleosome morphology and higher-level organisation; Heterochromatin and euchromatin, position effect variegation; Functional states of chromatin and alternation in chromatin organization. 3. CHROMOSOME ORGANIZATION: Metaphase chromosome; centromere and kinetochore, telomere and its maintenance; Telomeres and Aging.						

	 4. CHROMOSOME TERRITORIES: The Arrangement of Chromosomes in the Nucleus: Chromosomal domains (matrix, loop domains) and their functional significance; Dynamics of CT arrangements during postmitotic cell differentiation and in terminally differentiated cells. 5. CHROMOSOMAL ABNORMALITIES: Numerical (polyploidy, aneuploidy) and structural alterations (chromosomal rearrangements; deletion, duplication, inversion and translocation; structural abnormality: ring chromosomes and isochromosomes). Exercises: Telomere length analysis directly on chromosomes derived from primary cultured human skin fibroblasts and / or peripheral blood cells using quantitative fluorescence <i>in situ</i> hybridization, Q-PNA-FISH; application of molecular cytogenetic techniques (PCR, gel electrophoresis, immunofluorescence staining); optical fluorescence microscopy, image 						
	Seminars: Seminar is one of the <i>course</i> requirements. Students will have to prepare presentation on topics of the <i>original res</i> <i>paper related to the science unit they are studying</i> . The aim is to deve writing skills and presentation skills needed to effectively communicate purpose, scope, and conclusions of the project.						search ∍lop :e the
Format of instruction	 ☑ lectures ☑ seminars and workshops ☑ exercises □ on line in entirety ☑ partial e-learning □ field work 			 ☑ independent assignments ☑ multimedia ☑ laboratory □ work with mentor □ (other) 			
Student responsibilities							
Screening student	Class attendance	0,5	Research		Practical traini	ng	
proportion of ECTS credits for each activity so that the total number of ECTS credits is	Experimental work	0,5	Report		(Other)		
	Essay		Seminar essay	1,0	(Other)		
	Tests		Oral exam		(Other)		
value of the course)	Written exam		Project		(Other)		
Grading and evaluating student work in class and at the final exam	Research-based class seminar will be elevated. Students will have to prepare presentation showing background of the problem they are dealing with. The presentation will be scored according to the content of the presentation (key words, critical review of literature, presentation of scientific results), format, innovativeness and language competence as well.						
Required literature (available in the	Title				Number of copies in the library	Availabi other n	lity via nedia

library and via other media)	 Cooper, G.M., Hausman, R.E., 2015: Stanica- molekularni pristup. Šesto izdanje, Medicinska naklada, Zagreb 2015. Metode u molekularnoj biologiji, 2007. Andreja 					
	Abramovič Ristov (ur). Institut Ruđer Bošković.					
Optional literature (at the time of submission of study programme proposal)	 Molecular Biology of the Gene, Watson JD,Baker TA, Bell SP, Gann A, Levine M, Losick R, Pearson Education Inc., Benjamin Cummings, 2004. Practical in situ Hybridisation, Schwarcher T, Heslop Harrison P, Bios, Scientific Publisher Ltd. 2000. Plant Cytogenetics, Singh RJ, CRC Press London, 2003. Species Evolution: The Role of Chromsome Change, Max King, Cambridge University Press, 1995. Non radioactive in situ hybridisation application manual, Boehringer Mannheim, 1996. 					
Quality assurance methods that ensure the acquisition of exit competences	Student evaluation					
Other (as the proposer wishes to add)						