

1.1. Course description

NAME OF THE COURSE		Laboratory Course in Biophysics				
Code		Year of study	1 D			
Course teacher	Damir Kovačić Larisa Zoranić	Credits (ECTS)	4			
Associate teachers		Type of instruction (number of hours)	P	S	V	T
					40	
Status of the course	Obligatory	Percentage of application of e-learning				
COURSE DESCRIPTION						
Course objectives	The aim of the course is to introduce students to the biophysical, medical and biological methods of measurements and data analysis.					
Course enrolment requirements and entry competences required for the course	The learning outcomes of Bachelor programmes in physics, basic knowledge in molecular biology and biochemistry					
Learning outcomes expected at the level of the course (4 to 10 learning outcomes)	After completing the course, students will be able to: <ul style="list-style-type: none">do basic analysis of the morphology of neuronal cultures (density, directionality, velocity of growth)use numerical programs for the analysis of morphologymeasure and determine the concentration of peptidesmeasure and define the influence of peptides on the prokaryotic and eukaryotic cellsdo basic experiments on bacterial culturesunderstand basic atomic force microscopy (AFM) principlesanalyse AFM image with Gwyddion					
Course content broken down in detail by weekly class schedule (syllabus)	<div>1. Morphological analysis of neuronal cultures<ul style="list-style-type: none">Theoretical introduction - in vitro cultivation of neuronal cultures (4h)Image processing neuronal cultures of the software package ImageJ - remove the background, improve visual contrast, preparations for the Oval FFT (2h)Determination of the density and orientation of neuronal cultures (spiral and spinal ganglia) (4h)</div> <div>2. Antimicrobial peptides- measuring concentration and activity<ul style="list-style-type: none">Theoretical background – design and definition of biophysical characteristics by means of "on-line" tools. -(4h)Measure concentration of peptides by spectrophotometry - (2h)Measuring the inhibitory concentration of AMP - (2h)Measuring hemolytic activity - (2h)</div> <div>3 Electron microscopy<ul style="list-style-type: none">Principles of work of electron microscope - (2h)Sample preparation. Measurements - (6h)Data analysis - (2h)</div> <div>4. Atomic force microscopy (AFM)<ul style="list-style-type: none">Principles and modes of AFM imaging - (2h)</div>					

	<ul style="list-style-type: none"> • Cell preparation for AFM imaging. Cell imaging. - (6h) • Data analysis in Gwyddion software. - (2h)
Format of instruction	Laboratory experiments with an active participation of students and with professional guidance.
Student responsibilities	Student responsibilities are: to attend classes, to prepare and perform experiments, to write and presents reports.
Screening student work (name the proportion of ECTS credits for each activity so that the total number of ECTS credits is equal to the ECTS value of the course)	Classes – 1.5 Exams - 0.25 Written reports - 1.25 Final exam – 1
Grading and evaluating student work in class and at the final exam	During each term, the student's knowledge of the experiment is verified, while on each performed experiment students must write a report that will be evaluated. The final score is based on the knowledge shown during classes and exam,s and the written reports on conducted experiments.
Required literature (available in the library and via other media)	
Optional literature (at the time of submission of study programme proposal)	
Quality assurance methods that ensure the acquisition of exit competences	1. Analysis of the acquired learning outcomes at the end of the class, compared with the introductory work of students. 2. Monitoring the development of students in the subjects who followed the links with the success of the case 3. Other surveys of students
Other (as the proposer wishes to add)	